## Journal of Cardiovascular Magnetic Resonance

### Meeting abstract

# **147** <sup>31</sup>**P Cardiac spectroscopy at 3 T: T1 quantification** AbdEl-Monem M El-Sharkawy<sup>\*1</sup>, Michael Schär<sup>2</sup>, Ronald Ouwerkerk<sup>3</sup>, Robert G Weiss<sup>4</sup> and Paul A Bottomley<sup>3</sup>

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from 11<sup>th</sup> Annual SCMR Scientific Sessions Los Angeles, CA, USA. 1–3 February 2008

Published: 22 October 2008

Journal of Cardiovascular Magnetic Resonance 2008, 10(Suppl 1):A48 doi:10.1186/1532-429X-10-S1-A48

This abstract is available from: http://jcmr-online.com/content/10/S1/A48

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#### Introduction

Phosphorus (<sup>31</sup>P) MRS provides measures of the high energy metabolites, phosphocreatine (PCr) and adenosine triphosphate (ATP), in the heart. It permits the evaluation of ischemic changes during myocardial stress [1], and ATP turnover through the creatine-kinase reaction in the normal and failing human heart[2,3]. Recent cardiac <sup>31</sup>P MRS studies suggest higher signal-to-noise atio (SNR) at 3 T compared to 1.5 T in healthy subjects[4]. For accurate metabolite quantification, the longitudinal relaxation times (T1) are needed, and measuring these at 3 T is confounded by the combined effects of: (i) RF field uniformity with surface coil use; (ii) the available RF pulse power and its decrease with depth; and (iii) RF power deposition limits. While prior studies at 1.5 T used low-angle adiabatic (BIR4) pulses [2,3], at 3 T these are limited by low bandwidth and high power requirements. We show, using a Bloch equation analysis that such effects can significantly reduce the accuracy of T1 measurements at long adiabatic pulse lengths ( $\geq 10$  ms) for <sup>31</sup>P MRS, but that the problems are ameliorated by use of adiabatic half passage 90° (AHP) pulses.

The first aim of this work was to construct a high-SNR surface coil set for 3 T cardiac <sup>31</sup>P MRS that provides adequate adiabatic pulse power at the depth of the myocardium, while avoiding local power deposition problems. The second aim was to determine the T1 of PCr and  $\gamma$ -ATP in the human heart using a new, efficient dual repetition time (2TR) approach that minimizes T1 estimation errors at 3

T. The method is validated against the conventional saturation-recovery (SR) method.

#### Methods

A dual <sup>31</sup>P coil with 17-cm transmitter and 8-cm receiver set was designed and built to optimize the transmit RF field at a 10 cm depth with 4 kW transmit power. Coils were interfaced to a 3 T Achieva (Philips) broadband scanner. RF power deposition was computed and measured calorimetrically in phantoms to ensure safe performance. AHP pulses (10 ms) were tailored to achieve an excitation bandwidth  $\ge$  200 Hz for depths  $\le$  10 cm. Six healthy volunteers (4 M/2 F, 28  $\pm$  6 years) were positioned prone with the heart centered over the surface coils, as verified by scout-MRI. Localized second-order shimming[5] was performed, followed by cine-MRI to determine the period of least cardiac motion. The <sup>31</sup>P frequency was set between PCr and γ-ATP. Cardiac-gated one-dimensional chemical shift imaging was performed with TR = 2, 4, 12, 32 s with 24, 12, 4, and 2 averages, respectively (16 slices; 10 mm slice thickness; 2.5 kHz bandwidth).

T1 values for the human heart were determined from the signal  $S(TR) = M_0(1-exp(-TR/T1))$ , where  $M_0$  is the fully-relaxed magnetization, in three ways:

1) Conventional SR with a two-parameter least-squares fit;

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#### Figure I

A custom coil and protocol for human cardiac 3 T 31P MRS is used to measure the T1s of PCr and -ATP in the human heart via saturation recovery, and a new, efficient dual-TR approach that reduces bandwidth and power requirements. Left, end-systolic time-frame cardiac image showing coil center and 1DCSI slice locations. Right: Cardiac <sup>31</sup>P Spectrum (slice 7; TR = 12 s; 10 Hz filter; 13 min).

2) Point estimation (PE) with  $M_0 = S(TR = 32 \text{ s})$  as prior knowledge;

3) The new 2 TR method (with TRs of 2/12 s and 4/12 s)

In addition  $M_0$  was predicted  $(M_0^p)$  from the measured signal at the shorter TR and the estimated T1 from both of the 2 TR methods. Percentage errors were calculated as  $(M_0^{p-S}(TR = 32 \text{ s}))/S(TR = 32 \text{ s})*100\%$ .

#### Results

Cardiac spectra with PCr SNR  $\ge$  30 were acquired in all 6 volunteers (Figure 1). Conventional SR gave mean T1 values of 5.9 s for PCr and 3.1 s for  $\gamma$ -ATP (Table in Figure 2). Dual TR gave similar T1s in scan-times of 26 minutes, just

46% of the SR scan-time of 56 minutes, albeit with slightly larger errors.

#### Conclusion

Bandwidth and RF power limitations at 3 T necessitate significant modifications to <sup>31</sup>P MRS protocols as compared with 1.5 T. Our new dual-TR method provides fast cardiac <sup>31</sup>P spectra acquisition at 3 T, predicting the fully-relaxed magnetization within a 10% error compared with actual fully-relaxed values.

	Method	SR	PE TR=2	PE TR=4	2TR TR 2/12	2TR TR 4/12
PCr	T1 [s]	5.9 ±0.6	6.0±0.7	5.7 ±0.4	5.9 ±0.8	5.3 ±0.9
	M <sub>0</sub> % error	2 ±1			9 ±3	11 ±5
γ-ΑΤΡ	T1 [s]	3.1 ±0.6	3.2 ±0.5	3.3 ±0.6	2.7±0.5	2.8 ±0.9
	M <sub>0</sub> % error	4 ±2			9 ±4	10 ±3

#### Figure 2

Measured T1 values in 6 healthy volunteers (mean  $\pm$  stdev) for PCr and  $\gamma$ -ATP using the 3 methods as described in the text. Percentage error for the predicted fully relaxed magnetization are reported.

#### References

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